

LABORATORY REPORT

GROUP NUMBER: w3

EXPERIMENT NUMBER: 4

TITLE: Acid-Base Chemistry – Titration of an Amino Acid

DATE: March 3, 2000.

ROLE ASSIGNMENTS

<u>ROLE</u>	<u>GROUP MEMBER</u>
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ABSTRACT

A series of three acid-base titrations were conducted: HCl-NaOH, Na₂CO₃-HCl, and Alanine-NaOH; two pK_a's were determined for the alanine: pK_{a1}=2.43 with a confidence interval of 0 (this will be explained in the analysis) and pK_{a2} = 8.32, confidence interval of 0.28 as determined from the 95% Confidence test. Titration curves were constructed by titrating the sample with either NaOH or HCl, reading the pH meter at every interval and recording the data points in order to make semi logarithmic plots of pH versus moles of titrant. NaOH and HCl were restandardized and their concentrations were 1.00±0.05M. The manufacturer's nominal value for both were 1.000±0.005M. Though the manufacturer's values have a smaller uncertainty, they are essentially the same, our value of 1.00M lies within any possible confidence interval from the manufacturer that is centered around 1.000M. In order to perform a regression analysis, the original graphs of pH versus mol of titrant were modified so that a linear region could be determined. An altered form of the Henderson-Hasselbach equation was used (log([A⁻]/[HA]) versus pH in order to obtain a linear relationship for the determination of the pK_a's of the amino acid.

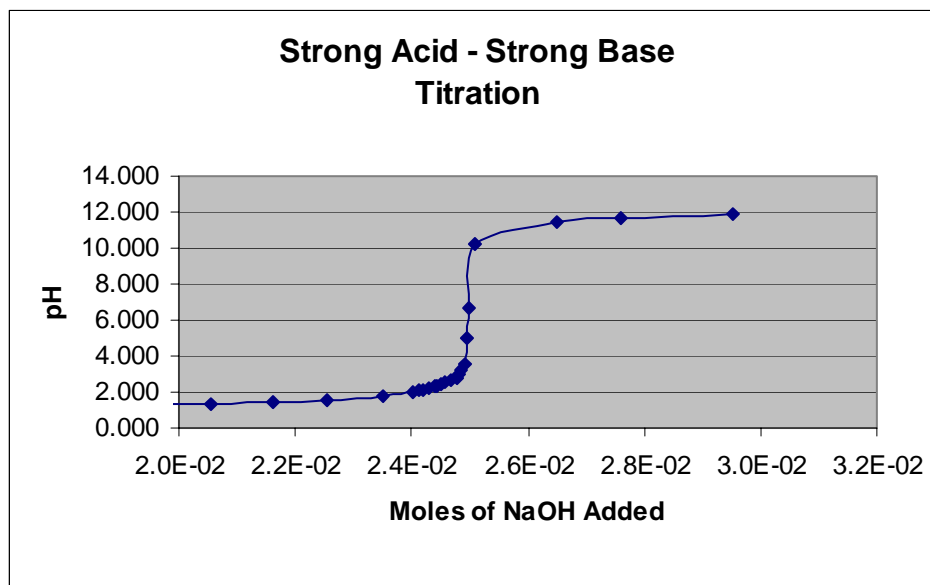
RESULTS

Table 1. Molarities of Secondary Acid and Base Standards

	Experimental Value (M)	Nominal Value (M)
HCl	1.00 +/- 0.05	1.000 +/- 0.005
NaOH	1.00 +/- 0.05	1.000 +/- 0.005

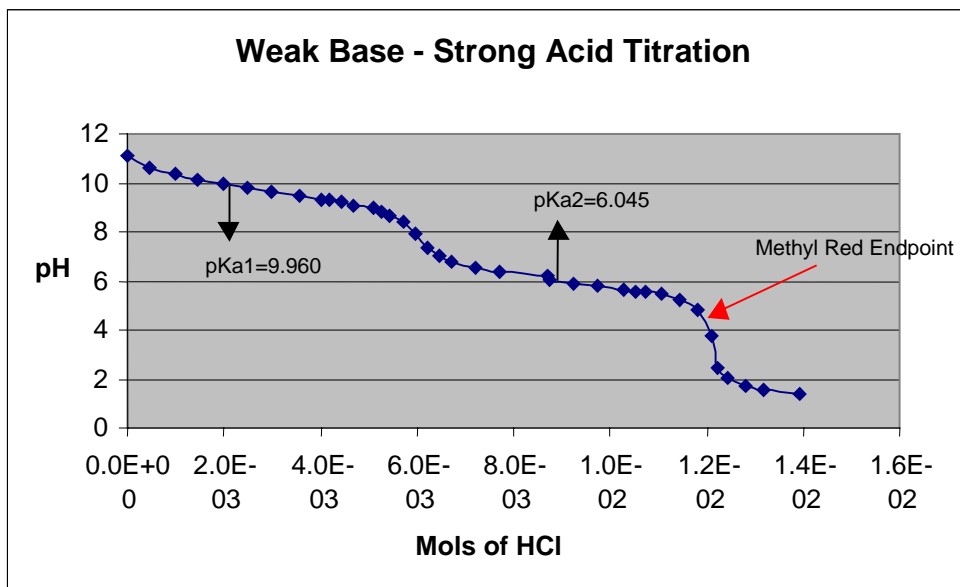
The experimental value obtained for the molarities of the secondary acid and base standards was 1.00 ± 0.05 M for both the HCl and NaOH. This value was obtained in two steps. First, an acid-base titration of HCL with NaOH revealed that 10.0 mL of NaOH was required to titrate 10.0 mL of HCl to the endpoint, which yields a ratio of 1.0. The only uncertainty in this step came from reading the pipettes (± 0.05 mL). After determining this ratio, the Na_2CO_3 primary standard was titrated to find the experimental values for the secondary standards (which are equal because of their 1-to-1 ratio). The calculation is as follows: $(1.3252 \text{ g Na}_2\text{CO}_3) \cdot (1 \text{ mol Na}_2\text{CO}_3 / 105.99 \text{ g Na}_2\text{CO}_3) \cdot (2 \text{ mol NaOH} / 1 \text{ mol Na}_2\text{CO}_3) (1 / 0.2491 \text{ L NaOH}) = 1.00 \pm 0.05$ M. The manufacturer's nominal value was 1.000 ± 0.005 M.

Figure 1. HCl-NaOH Titration Curve



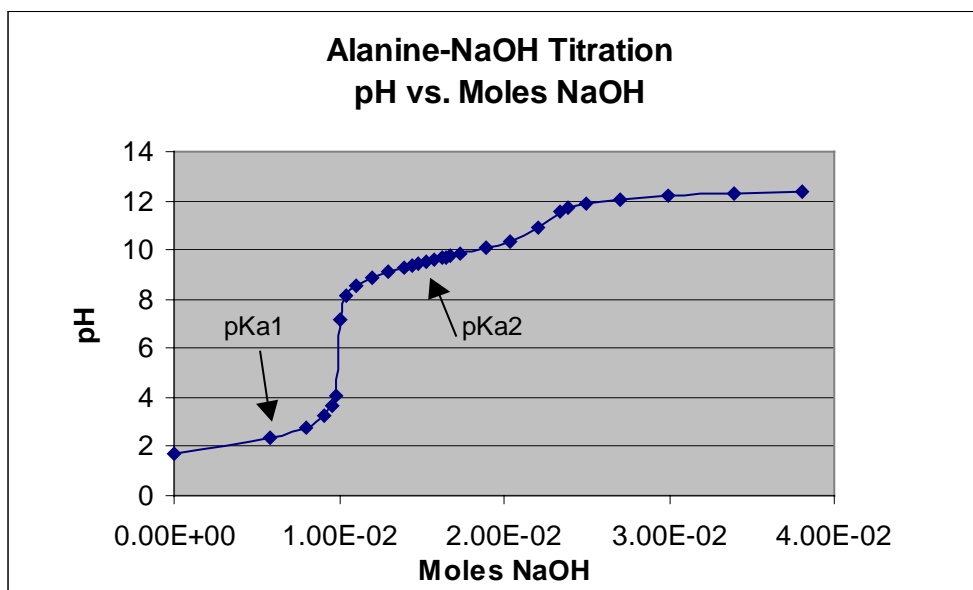
A titration of HCl with NaOH revealed that 2.5×10^{-2} moles of HCl was neutralized after 2.5×10^{-2} moles of NaOH has been added. This value corresponds exactly to the end point as determined by the phenolphthalein indicator.

Figure 2. Na_2CO_3 -HCl Titration Curve



For the titration of Na_2CO_3 with HCl, $\text{pK}_{a1} = 9.960$ and $\text{pK}_{a2} = 6.045$. The red arrow indicates the value of the Methyl Red endpoint, which occurs between pH 4 and 5.

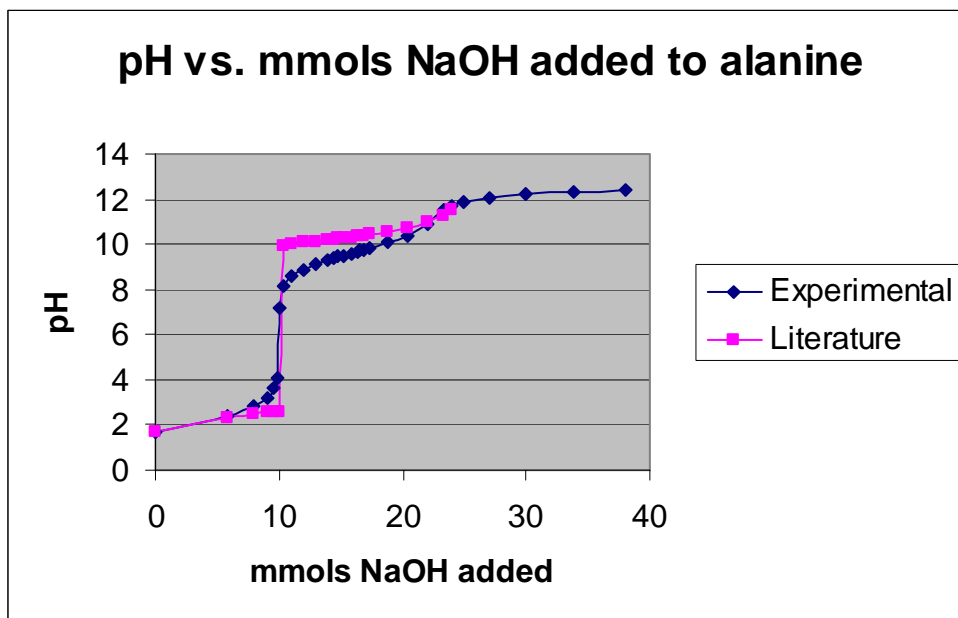
Figure 3. Alanine-NaOH Titration Curve



Arrows indicate the pKa values of the alanine amino acid titration with NaOH using a methyl red indicator, $\text{pK}_{a1} = 2.43$ confidence interval of 0 (explanation in analysis) and $\text{pK}_{a2} = 8.32$ with confidence interval of 0.28. In order to perform a regression analysis, the original graphs of pH versus mol of titrant were modified so that a linear region could be determined. An altered form of the Henderson-Hasselbach

equation was used ($\log([A^-]/[HA])$) versus pH in order to obtain a linear relationship for the determination of the pK_a 's of the amino acid. The plot thus constructed (Y vs. pH) follows in Figure 5. Several regression analyses were performed to determine the linear regions in the graph. This is done by applying the 95% confidence interval test for the determination of the best confidence interval.

Figure 4: Titration of Alanine with NaOH: pH vs. mmols NaOH added (comparison of experimental and literature values)



Experimental alanine titration curve against literature titration curve is plotted above. The literature titration curve was plotted using the two given pK_a values of 2.35 and 9.69 as a basis. The graph was constructed keeping the x values (mmols of NaOH added) at the same value as the experimental while the y-values (pH) was calculated using the given pK_a values. Points of $n=1-6$ (where n indicates the nth point) were plotted using pK_a of 2.35, and that of 7 to 20 were plotted using pK_a of 9.69. The literature titration curve has sharp angles because pK_a values can only determine the shape of the graph around where the pK_a would be taken, or the areas roughly in the center between two end points. pK_a values cannot be used to construct a curve that represents the area around the end points.

It can be seen from this graph, that the titration curve around the first pK_a is approximately equal in the two curves, that of the second pK_a is very different. The slope around the second pK_a is much higher (steeper) in the experimental than in the literature. To quantitatively account for this difference, the Y vs. pH plot for the two values are drawn and presented in Figure 5.

approximately 8pH units over a very small volume change. Being that there is only one pKa for both graphs, they are both titrations of strong monoprotic acids with one to one molar ratios with their respective base.

The Na₂CO₃ titration graph in the appendix of the lab manual¹ is a titration of carbonic acid with NaOH. In the case of this lab however, Na₂CO₃, a weak base, is titrated by HCl. The graphs however both yield the same information, when the carbonic acid is being titrated by a strong base the graph starts at an acidic pH (around 4) and raises to a basic value of about 13. In the lab, the Na₂CO₃ was titrated by HCl and because of this the graph is reversed form that found in the appendix, meaning that it begins at a basic value (13pH) and falls to an acidic pH of 1.5. The pKa 's for the two graphs are reversed, pKa₁ for one graph is pKa₂ for the other. The appendix graph has pKa₁ and pKa₂ values of 6.38 and 10.38 respectively. The lab data graph yields a pKa₁ of 9.960 and pKa₂ of 6.045. Understanding the reversed relationship of the two graphs, the pKa₁ value obtained in lab deviated for the book value by 5.25%. Similarly, the pKa₂ value obtained in lab differs from the book value by 3.49%. The methyl red endpoint signifies the point at which all of the Na₂CO₃ has become H₂CO₃. Every point before with the exception of the initial start of the titration has been an acid base mixture.

The pK_a values of alanine were determined to be pK_{a1}=2.43 with a confidence interval of 0 and pK_{a2} = 8.32, confidence interval of 0.28 as determined from the 95% Confidence test. Literature values for alanine's pK_{a1} and pK_{a2} are 2.35 and 9.69, respectively. Experimental values are 2.43 and 8.32, respectively.² The percent error's are 3.29% for pK_{a1} and 14.14% for pK_{a2}. The experiment pK_{a1} in comparison to the literature value is very similar, however the second pK_a values do not show much resemblance to the literature. This will be discussed subsequently.

A regression analysis was conducted on the first three points of the graph yielding a confidence interval of 8.14. This confidence interval was not suitable because the last point that was included for the test does not even lie in the linear region of the curve, therefore cannot be included. Subsequently, a regression analysis was done using the only two points in the linear region of the curve. This was technically not possible to do since regressions cannot be done on two points. However, there were no other points in the linear region; therefore, a true regression analysis could not have been accomplished, thus a 0 confidence interval was determined. In order to obtain an accurate confidence interval for this first region of the curve, more points should have been recorded along with their pH values.

In the second linear region of Figure 5, a pK_{a2}=8.32 was yielded. The confidence interval for this pK_{a2} is 0.28. The same method was utilized—testing different points on Figure 5

¹ Litt, Mitchell. Acid-Base Titrations: Titration of an Amino Acid. *Bioengineering Laboratory Manual*. p. 14, 2000

² Litt, Mitchell. Acid-Base Titrations: Titration of an Amino Acid. *Bioengineering Laboratory Manual*. p. 12, 2000

in order to determine the best confidence interval. In order to obtain this confidence interval points (8.90, 0.31 through 11.51, 1.60) were tested. Other confidence intervals included: (8.90, 0.37 through 11.60, 1.80 yielded 0.57 confidence; 9.30, 0.52 through 10.37, 1.03 yielded 0.34 confidence); however, as shown one can see that the region between points 8.90, 0.31 through 11.51, 1.60 yield the finest confidence interval.

There are a number of errors that must be accounted. First, the pH meter has difficulty reading pH's in this portion of the pH scale. Basic solutions tend to dissolve the glass, thus the pH meter gives very unstable reading around any pH higher than 9. This was seen when the calibration of the pH meter was done in the first part of the experiment when the pH of the buffer was 10. The meter had a difficult time stabilizing in this area. Such error is only minimal, however. When the pH meter was used to read the pH 10 buffer, a value of 10.03 was read instead of 10.00. This amounts to 0.3% error.

Second, there is an uncertainty on the 50-mL pipette, corresponding to an error of ± 0.02 mL for each reading.

Neither of these errors, however, can account for the grave experimental percentage error of 14.14% for pK_{a2} . The most probable explanation for this error is systematic human error, arising from the mistake of not waiting long enough for the pH meter to stabilize. When the pH meter was used to read the pH 10 buffer (giving a value of 10.03 instead of 10.00, as described above), the experimenter waited approximately 30 seconds, during which time the value slowly climbed up to 10. During the course of the actual experiment, however, the experimenter waited an approximately 5 seconds only due to time constraints. As such, it is probable that the pH meter was not able to stabilize and reach its final reading before the reading was taken. This would account for the readings which are consistently below what the literature value would dictate.

In conclusion, three acid-base titrations were conducted and both pK_a values for alanine ($pK_{a1}=2.43$ with a confidence interval of 0 and $pK_{a2} = 8.32$, confidence interval of 0.28 as determined from the 95% Confidence test) were determine using a liner regression method and modifications of the Henderson-Hasselbach equation. Restandardazations of both the HCl and the NaOH were done yielding concentrations of $1.00+0.05M$ for both and were then compared to the nominal manufacturer values of $1.000\pm 0.005M$ for both the acid and the base. Our value of 1.00 M lies within any possible confidence interval from the manufacturer which is centered around 1.000 M, thereby definition is not significantly different.

Both educational goals were met. Quantitative measurements of pH using modern instrumentation (pH meter) and classical acid-base indicators (methyl red, phenol red, and phenolphthalein) were used in each titration. Acid-base properties of a biochemical electrolyte, alanine, were studied by use of titration techniques. Additionally, measurements of important equilibrium properties of the reactions involved were studied and analyzed by the techniques developed in the experiment.

REFERENCES

Litt, Mitchell. Acid-Base Titrations: Titration of an Amino Acid. *Bioengineering Laboratory Manual*. p. 12, 2000